



# *Enteric gram-negative bacterial infections of the Gastrointestinal tract 2*

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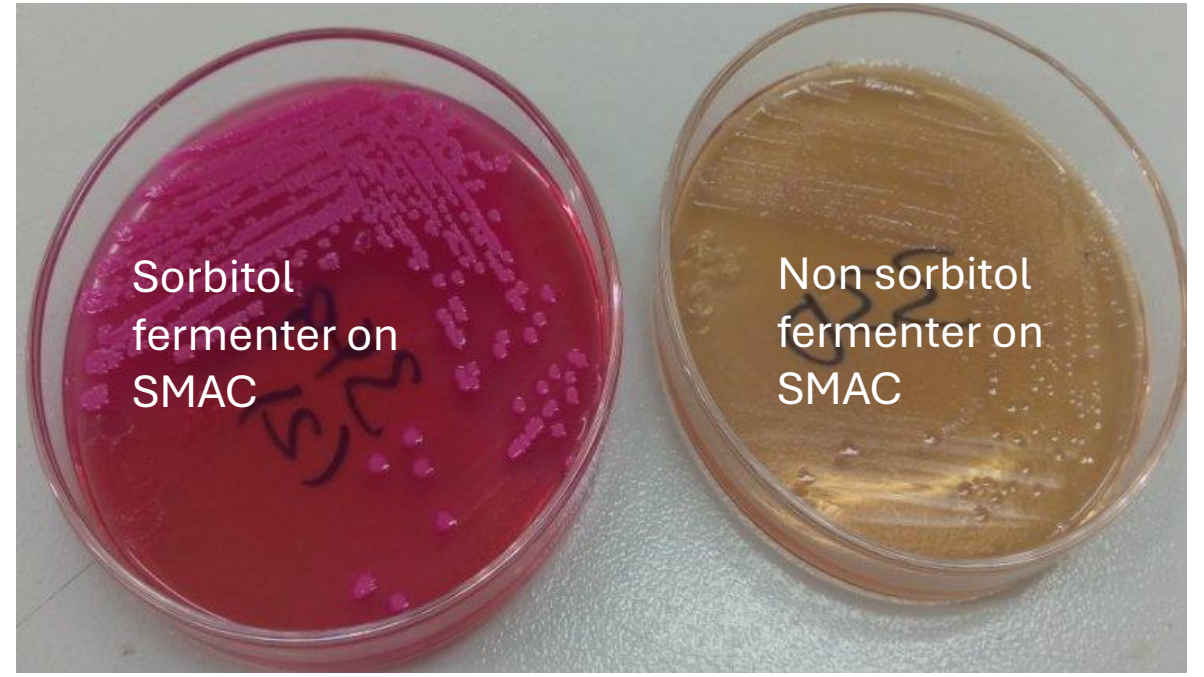
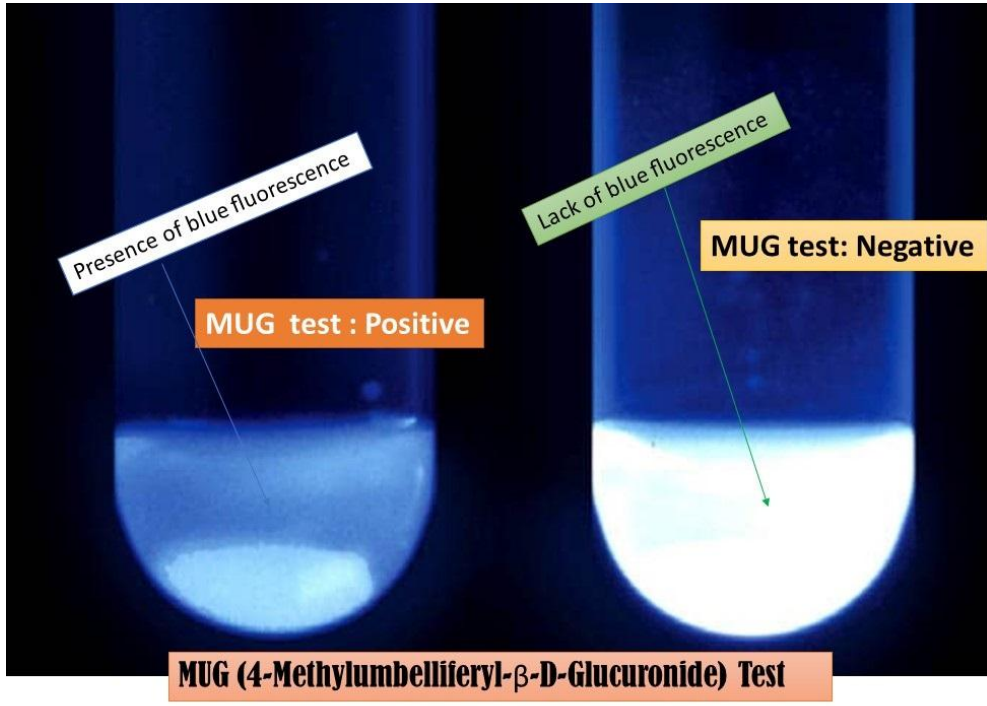
School of Medicine

# Diagnosis of Diarrheagenic *E. coli*: EPEC and ETEC

- **EPEC** strains are identified by **O antigen** and occasionally by **H antigen typing**, and a two-stage infection model using HEp-2 or HeLa cells also can be performed for EPEC (in reference labs).
- Assays for **ETEC heat-labile toxin (LT)**, such as cell cultures, immunologic assays and gene detection, are used in reference labs.
- The plasmids carrying the genes for enterotoxins may also carry genes for the **colonization factor antigens (CFAs)** that facilitate attachment of *E. coli* strains to intestinal epithelium.

# Diagnosis of STEC, EIEC, and EAEC

- **STEC O157:H7** does not use **sorbitol**, unlike most other *E. coli*, and is **negative (clear colonies)** on **sorbitol MacConkey agar (SMAC)**.
- O157:H7 strains also are negative for  **$\beta$ -glucuronidase** using the substrate **4-methylumbelliferyl- $\beta$ -glucuronide (MUG test)**.
- **Specific antisera** are used to identify the **O157:H7 strains**, and enzyme immunoassays (EIAs) for Shiga toxins detection.
- Other tests for STEC: cell culture cytotoxin testing using Vero cells and **polymerase chain reaction for direct detection of toxin genes** directly from stool samples.



- **EIEC** strains are similar to *Shigella* and are **non-lactose or late lactose fermenters** and are **nonmotile**.
- **EAEC** can be suspected clinically but requires confirmation by **tissue culture adhesion assays**, which are **not readily available in most clinical laboratories**.

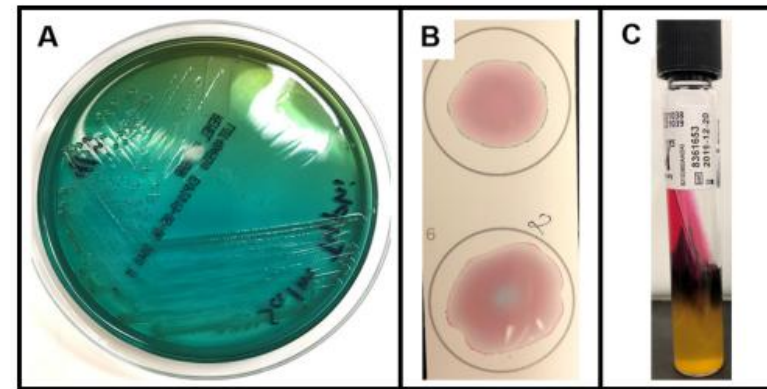
# Treatment and Prevention of Diarrheagenic *E. coli*

- The duration of **EPEC diarrhea** can be shortened and the chronic diarrhea cured by **antibiotic treatment**.
- When **ETEC diarrhea** develops, **antibiotic treatment effectively shortens the duration of disease**.
- For **traveler's diarrhea**, antimicrobial prophylaxis can be effective but may result in increased **antibiotic resistance** and probably should not be uniformly recommended.

- It is widely recommended that **caution be observed in regard to food and drink** in areas where environmental sanitation is poor.
- **Early and brief treatment**, for example with **ciprofloxacin** or **trimethoprim-sulfamethoxazole**, should be substituted for prophylaxis of traveller's diarrhea.
- Many cases of **hemorrhagic colitis** and its associated complications can be prevented by **thoroughly cooking ground beef** and by **avoiding unpasteurized products such as apple cider**.

# *Salmonella* microbiological and biochemical features

- **Salmonellae are non-spore-forming, facultative anaerobic, Gram-negative bacilli** that vary in length.
- Most isolates are **motile with peritrichous flagella**.
- Salmonellae grow readily on **simple agar media**.
- They are able to utilize **citrate as a sole carbon source** and **lysine as a nitrogen source**, almost never ferment **lactose or sucrose**.
- Salmonellae are **oxidase negative**.
- They form **acid and sometimes gas** from fermentation of **glucose and mannose**.
- They usually produce **H<sub>2</sub>S**.



**FIG 1** Growth and biochemical characteristics of *Salmonella enterica* subsp. *enterica* serovar Typhi. (A) Growth on Hektoen enteric agar (non-lactose fermenter, slight H<sub>2</sub>S production). (B) Serological reactivity to Remel Wellcolex Color *Salmonella* test reagents 1 (top: green agglutination with red background = serogroup D) and 2 (bottom: red agglutination with blue background = Vi antigen), consistent with *S. Typhi*. (C) Growth on a triple sugar iron agar slant (K/A with slight H<sub>2</sub>S production).

<https://journals.asm.org/doi/pdf/10.1128/jcm.01359-20>

**TABLE 15-4 Clinical Diseases Induced by Salmonellae**

	Enteric Fevers	Septicemias	Enterocolitis
Incubation period	7–20 days	Variable	8–48 hours
Onset	Insidious	Abrupt	Abrupt
Fever	Gradual; then high plateau with “typhoidal” state	Rapid rise; then spiking “septic” temperature	Usually low
Duration of disease	Several weeks	Variable	2–5 days
Gastrointestinal symptoms	Often early constipation; later, bloody diarrhea	Often none	Nausea, vomiting, diarrhea at onset
Blood culture results	Positive in first to second weeks of disease	Positive during high fever	Negative
Stool culture results	Positive from second week on; negative earlier in disease	Infrequently positive	Positive soon after onset

# Diagnostic Specimens

- **Freshly passed stool** is the preferred specimen for diagnosis of **non-typhoidal *Salmonella***.
- Specimens collected during the **early stages** of enteric illness have the highest yield.
- Collection of **multiple stool specimens** may enhance recovery of *Salmonella* and other enteric pathogens (eg, *Shigella*)..
- For definitive diagnosis of **enteric fever**, ***Salmonella Typhi*** or ***Salmonella Paratyphi*** must be isolated in culture.

- Appropriate specimens for enteric fever include **blood (most commonly used), bone marrow, other sterile sites, urine, or intestinal secretions.**
- Blood cultures are often positive in the **first week** of enteric fever and septicemia.
- **Bone marrow cultures** have the highest sensitivity, **80% to 95%**, but are clinically less practical.
- Urine culture may become positive after the **second week** of illness.

## Culture and Laboratory Identification

- Specimens may be cultured on **differential media**, including **EMB (eosin methylene blue)**, **MacConkey**, or **deoxycholate medium**, which permit rapid detection of **lactose non-fermenters**.
- **Bismuth sulfite medium** permits rapid detection of salmonellae, which form **black colonies because of H<sub>2</sub>S production**.
- **Selective media** include **salmonella-shigella agar**, **Hektoen enteric agar**, **xylose-lysine desoxycholate agar**, and **desoxycholate-citrate agar**.
- These selective media favor growth of salmonellae and shigellae over other **Enterobacteriaceae**.

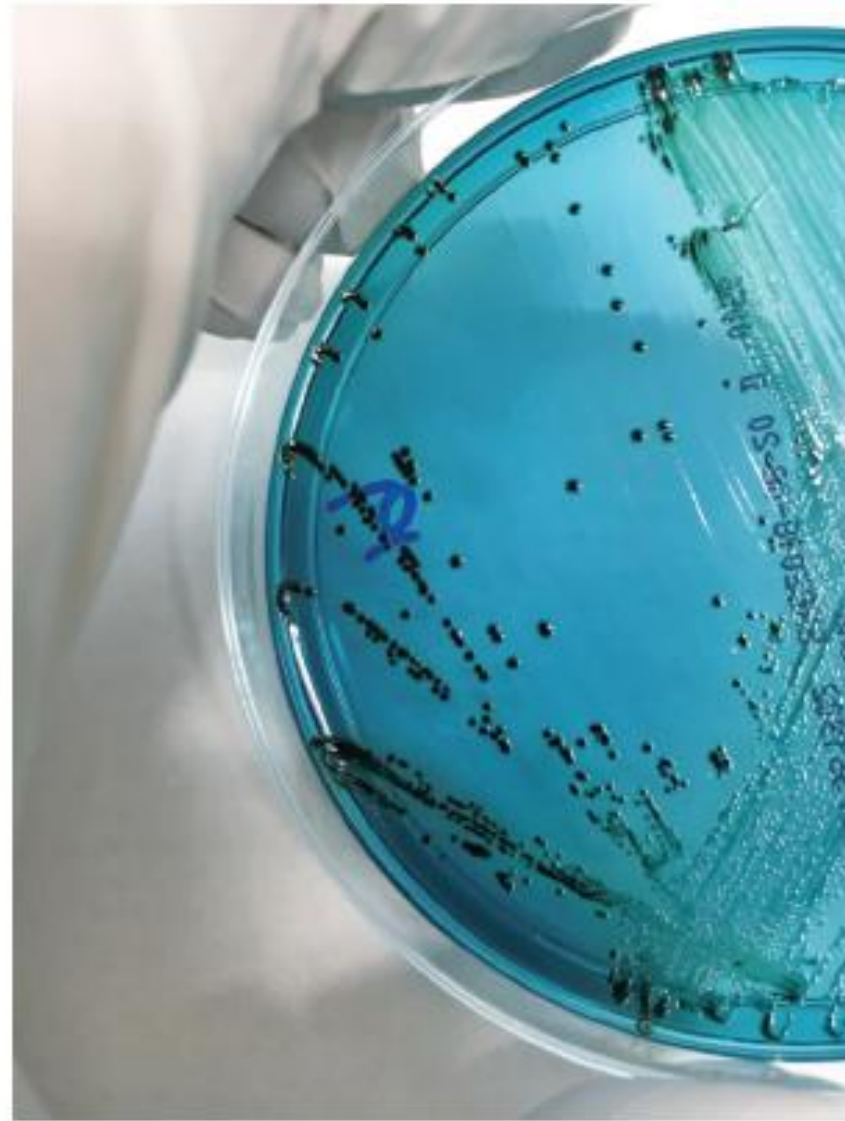


FIG. 1. *Salmonella enterica*. Note the black center of the transparent colonies indicating H<sub>2</sub>S production in the absence of carbohydrate utilization. All serotypes of *Salmonella* have this appearance on HE agar except for serotype Typhi, which is a weak H<sub>2</sub>S producer, and rare strains of lactose-fermenting *Salmonella*.

<https://asm.org/asm/media/protocol-images/hektoen-enteric-agar-protocol.pdf?ext=.pdf>



A



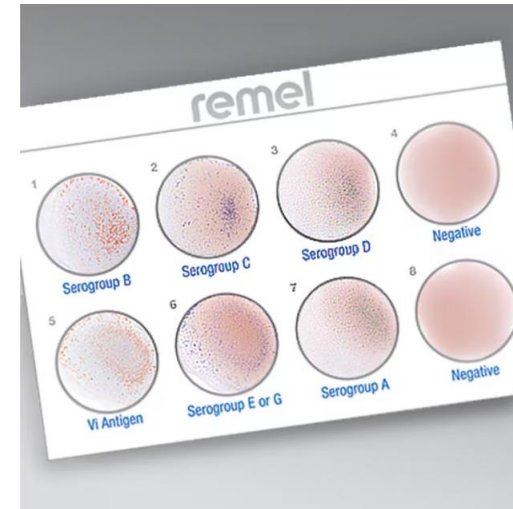
B

**FIGURE 15-4** **A:** *Salmonella* species on TSI agar slant. **B:** *Salmonella* species on HE agar. *Salmonella* spp. do not ferment the carbohydrates present in the HE agar; however, the organism produces  $H_2S$ , and the ferric ammonium citrate in the HE agar result in the *Salmonella* colonies to appear black. On TSI agar slant, *Salmonella* spp. ferment glucose, but not lactose; they produce  $H_2S$  and gas [K/A,G  $H_2S^+$ ]. (Courtesy of S. Riedel.)

- **Chromogenic agars** specifically for Salmonella recovery are also available.
- **Enrichment cultures:** Stool specimens can also be placed into **selenite F or tetrathionate broth**, which inhibit normal intestinal bacteria and permit multiplication of salmonellae.
- Suspect colonies from solid media are identified by **biochemical reaction patterns** and **slide agglutination tests with specific sera**.

## Serology and Molecular Diagnosis

- Serologic techniques are used to identify unknown cultures with known sera, but antibody testing is **not very useful** for diagnosis of *Salmonella* infections.
- In the **agglutination test**, known sera and unknown culture are mixed on a slide, and clumping can be observed within a few minutes.
- Commercial kits are available to agglutinate and serogroup salmonellae by **O antigens: A, B, C1, C2, D, and E.**



Wellcolex™ Color Salmonella Rapid Latex Agglutination Test Kit

**TABLE 15-3** Representative Antigenic Formulas of *Salmonellae*

O Group	Serotype	Antigenic Formula*
D	<i>Salmonella</i> Typhi	9, 12 (Vi):d:—
A	<i>Salmonella</i> Paratyphi A	1, 2, 12:a—
C <sub>1</sub>	<i>Salmonella</i> Choleraesuis	6, 7:c:1,5
B	<i>Salmonella</i> Typhimurium	1, 4, 5, 12:i:1, 2
D	<i>Salmonella</i> Enteritidis	1, 9, 12:g, m:—

\*O antigens: boldface numerals.

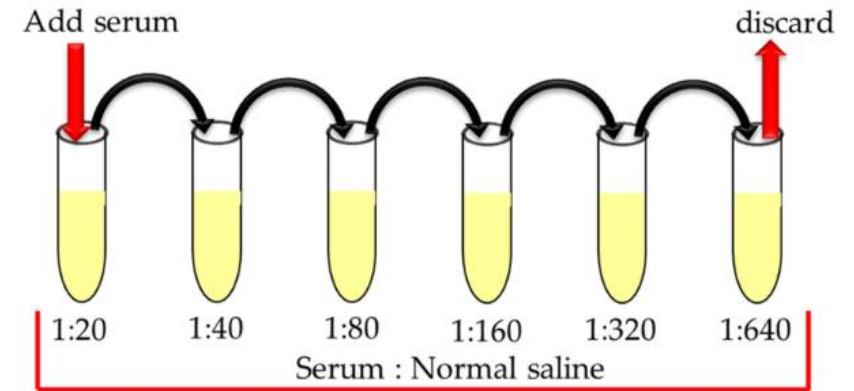
(Vi): Vi antigen if present.

Phase 1 H antigen: lowercase letter.

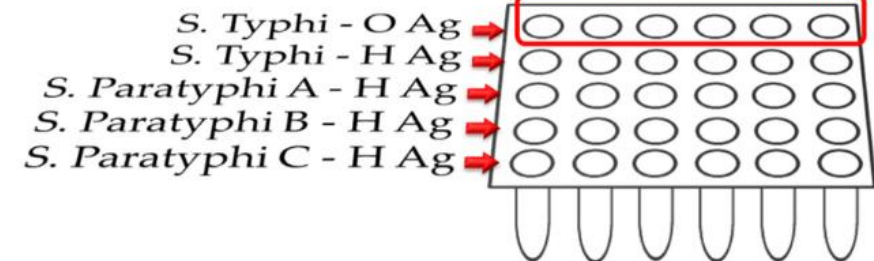
Phase 2 H antigen: numeral.

- **Tube dilution agglutination test (the Widal test)** detects antibodies against **O and H antigens** in suspected **S. Typhi** infection.
- At least two serum specimens, obtained **7–10 days** apart, are needed to prove a rise in antibody titer.
- False-positive and false-negative results occur, so serologic tests cannot be relied upon to establish a definitive diagnosis of typhoid fever.
- A single titer against the O antigen of greater than 1:320 and against the H antigen of greater than 1:640 is considered positive.
- High titer of antibody to the Vi antigen occurs in some carriers.
- Several commercial **NAATs** are available for direct detection of salmonellae in fecal samples of patients with **acute diarrhea**.

### 1. Serial dilution



### 2. Add antigens



### 3. Mix properly, cover and incubate at 37° C overnight




# Immunity

- Infections with *Salmonella* Typhi or *Salmonella* Paratyphi usually confer a certain degree of immunity.
  - While reinfection may occur, it is often milder than the initial infection.
  - Circulating antibodies to O and Vi are related to resistance to infection and disease, however, relapses may occur in 2–3 weeks.
  - Children, with sickle cell disease or sickle cell trait are more susceptible to *Salmonella* infections particularly to bacteremia and its complications (eg, osteomyelitis).
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# Treatment of *Salmonella* Infections

- **Non-typhoidal *Salmonella* gastroenteritis** is typically self-limited; antimicrobial therapy is usually **not necessary and not recommended**.
  - In severe diarrhea, **replacement of fluids and electrolytes is essential**.
  - Antimicrobial treatment of *Salmonella* gastroenteritis should be considered in **neonates**, immunosuppressed patients, and patients older than **50 years** with suspected or confirmed vascular disease.
  - For susceptible organisms, oral therapy with **amoxicillin, trimethoprim-sulfamethoxazole, or a fluoroquinolone** is appropriate.
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- **Uncomplicated enteric fever** can be managed with oral **azithromycin, 1 g once**, followed by **500 mg daily for 7 days**.
  - Patients with complications should be hospitalized and treated with a parenteral **third-generation cephalosporin** or **fluoroquinolone** for at least **10 days**.
  - In chronic carriers, ampicillin combined with cholecystectomy can be therapeutic.
  - Non-typhoidal *Salmonella* bacteremia should be empirically treated with a **third-generation cephalosporin**, such as **ceftriaxone**, and a **fluoroquinolone** until antimicrobial susceptibility testing results are available.
  - Endovascular infection (eg, infected aneurysm) should be treated with intravenous **ceftriaxone, ampicillin, or a fluoroquinolone** for **6 weeks**, followed by oral therapy; early surgical resection of an infected aneurysm is recommended.
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# Shigellae microbiological, biochemical, and identification Features

- Shigellae are **slender, nonmotile Gram-negative rods; coccobacillary forms** occur in young cultures.
- Shigellae are **facultative anaerobes** but grow best **aerobically**.
- All strains of ***Shigella* species ferment glucose**.
- With the exception of ***Shigella sonnei***, they **do not ferment lactose**.



Pleural Empyema Due to *Salmonella typhi* - Scientific Figure on ResearchGate. Available from: [https://www.researchgate.net/figure/Non-lactose-fermenting-colonies-on-MacConkey-agar\\_fig2\\_233875609](https://www.researchgate.net/figure/Non-lactose-fermenting-colonies-on-MacConkey-agar_fig2_233875609)

- Shigellae form **acid from carbohydrates** but **rarely produce gas**.
- They may also be divided into those organisms that **ferment mannitol** and those that **do not**.
- The pathogenic species are ***S. sonnei*, *S. flexneri*, *S. dysenteriae*, and *S. boydii***.

**TABLE 15-2** Pathogenic *Shigella* Species

Present Designation	Group and Type	Mannitol	Ornithine Decarboxylase
<i>Shigella dysenteriae</i>	A	–	–
<i>Shigella flexneri</i>	B	+	–
<i>Shigella boydii</i>	C	+	–
<i>Shigella sonnei</i>	D	+	+

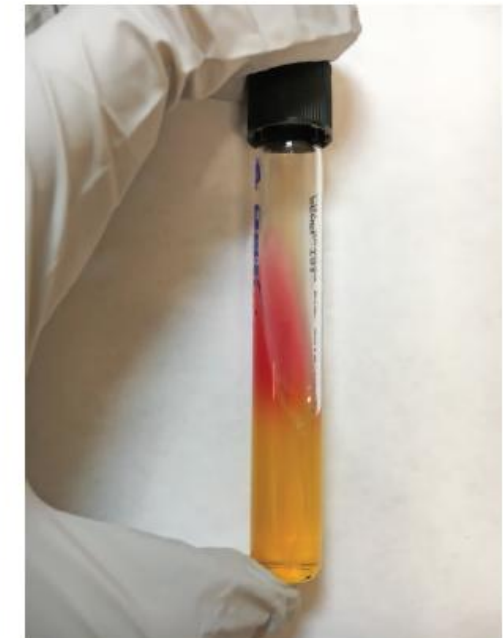
# Diagnostic Laboratory Tests and Identification

- For optimal organism recovery, **fecal specimens** should be collected during the **early stages** of the illness.
- Specimens include **fresh stool, mucus flecks, and rectal swabs** for culture.
- While whole stool is usually the preferred specimen for laboratory workup of diarrhea, **rectal swabs with visible fecal staining** may be the preferred specimen for the isolation of shigellae.
- The materials are streaked on **differential media** such as **MacConkey or EMB agar** and on **selective media** such as **Hektoen Enteric agar** or **xylose-lysine-deoxycholate agar**.
- These selective media suppress other **Enterobacteriaceae** and **Gram-positive organisms**.

- **Colorless (lactose-negative) colonies** are inoculated into a **TSI agar slant**.
- **Nonmotile organisms** that fail to produce **H<sub>2</sub>S**, that produce **acid but not gas in the butt and an alkaline slant in TSI agar medium**, should be subjected to **slide agglutination by specific *Shigella* antisera**.
- It should be noted that ***Shigella* species and *E. coli* cannot be reliably differentiated by MALDI-TOF MS**.
- There are several commercial **NAATs** that directly detect shigellae in fecal samples along with some of the other major enteric pathogens.
- **Serology is not used to diagnose *Shigella* infections**.



A



B

**FIGURE 15-3** **A:** *Shigella* species on Hektoen Enteric (HE) agar. **B:** *Shigella* species on TSI agar slant. *Shigella* spp. do not ferment lactose, salicin, or sucrose, and therefore appear as translucent, colorless colonies on HE agar. *Shigella* spp. ferment glucose, but not lactose and sucrose present in the TSI agar slant. Therefore, they produce an alkaline over acid (K/A) reaction and do not produce H<sub>2</sub>S or gas. (Courtesy of S. Riedel.)

# Treatment of Shigellosis

- In general, **shigellosis is a self-limited illness**, and many patients recover without treatment within **5–7 days**.
- Mortality is generally low in shigellosis, except in **malnourished children, infants, and elderly patients**.
- Severe dehydration, **febrile seizures, septicemia, and pneumonia** are potential complications of severe shigellosis.
- In general, **oral fluid replacement** is considered to be sufficient for treatment of uncomplicated shigellosis.
- In high-risk patient populations, **intravenous fluid replacement** may be required.
- **Antidiarrheal medications** such as **loperamide and opioids** should be avoided in **Shigella dysentery**, as such medications may worsen the symptoms of the illness.

- **Antibiotic treatment is recommended** for the treatment of **severe infections** and to prevent secondary spread among people living in closed quarters or during outbreaks.
- Because of widespread resistance, **trimethoprim-sulfamethoxazole and ampicillin are no longer recommended as first-line agents** for treatment of shigellosis.
- **Ciprofloxacin and ceftriaxone are effective antibiotics of choice.**
- **Ceftriaxone** is commonly used for treatment of **children with shigellosis.**
- **Azithromycin** has been shown as a useful antibiotic for treatment of **antibiotic-resistant Shigella infections** in adults and children.

- Riedel, S., et al. (Eds.). (2026). *Jawetz, Melnick, & Adelberg's Medical Microbiology* (29th ed.). McGraw Hill –chapter 15
- Ryan, K. J. (Ed.). (2022). *Sherris & Ryan's Medical Microbiology* (8th ed.). McGraw Hill- chapter 33